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and mean P-N value among positive samples. When these hepatitis cases were analyzed by subgroup, the highest prevalence (47%) of antibody to the SMSV pool was in the cases of Non-A-E hepatitis associated with transfusion or dialysis. Cases of Non-A-E hepatitis with an unknown exposure source had an estimated prevalence (19%) similar to that (21%) of the high ALT donors and cases of HBV or HCV hepatitis had an estimated prevalence (10%) similar to that of healthy donors. The increase of anti-SMSV pool prevalence along the axis "Normal donor-high ALT donor-Non-A-E hepatitis case associated with transfusion or dialysis" was highly significant (P < 0.001, χ^2 for the trend). The mean P-N value of the positive samples from the Non-A-E hepatitis group was lower than that of the other groups studied (P > 0.05 for these comparisons). The patterns of significant differences in estimated prevalence were similar and statistically significant for a range of antibody assay cut-points, up to 0.200, although estimated prevalence declined as the cut-point increased.

Detection of Vesivirus RNA in Human Sera

Total RNA was extracted from 30 donor sera and tested by RT-PCR 1. The laboratory performing this testing had no prior experience with *Vesivirus* genomes in the facility. RT-PCR 1 generated a *Vesivirus* amplicon in one of these samples that was confirmed by dot-blotting (Fig. 2) and sequencing.

To reduce the possibility that a positive result for serum Vesivirus genome detection was generated by laboratory contamination at the first facility, a further 82 donor samples were tested in a second laboratory routinely performing RT-PCR and that had prior experience with Vesivirus RNA, but using newly designed primers to amplify a genomic region not previously amplified (RT-PCR 2). Ten of these 82 samples yielded amplicons of the expected size (Fig. 3).

Ten (11%) amplicons came from 91 high ALT donor samples and 1 (4.8%) amplicon from 21 Normal donor

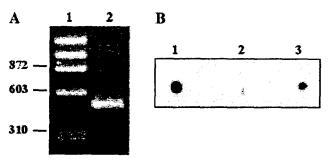


Fig. 2. RT-PCR 1 product and confirmatory dot blot. RT-PCR 1 testing of blood donor sera revealed an amplicon of the expected size for Vesivirus RNA (A) and dot blot probing of total RNA from the same serum provided evidence of Vesivirus viremia (B). In (A), Lane 1: molecular weight ladder; lane 2: RT-PCR 1 amplicon. In (B), dot 1 is 20 ng of RNA extracted from CsCl-banded SMSV-5, dot 2 is 50 ng of total RNA from mouse embryo, and dot 3 is 50 ng of total RNA extracted from the donor serum sample.

samples $(P=0.64, \text{ Yates' corrected } \chi^2)$. Five (15%) of the amplicons were from 34 sera that scored EIA-positive and 6 (7.7%) from 78 sera that scored EIA-negative $(P=0.42, \text{ Yates' corrected } \chi^2)$.

Sequence Comparisons of the RT-PCR Amplicons

Six amplicons from the RT-PCR 2 (polymerase region) sample set and the one from the RT-PCR 1 (capsid region) sample set were successfully sequenced. Amplicons not successfully sequenced yielded a band too faint for successfull direct sequencing and were not successfully cloned. Five of the six polymerase region amplicons were distinct from each other but closely related (1–6% divergence). The sixth amplicon, study number N104, was distinct from the other five polymerase region amplicons (24–38% divergence from N104). When the five similar polymerase region amplicons were compared with GenBank entries spanning the amplification region of RT-PCR 2, the two best matches were with primate Vesivirus Pan-1 (88–96% identity) and SMSV-6 (88–94%) and lower match was with SMSV-5 (84–86%).

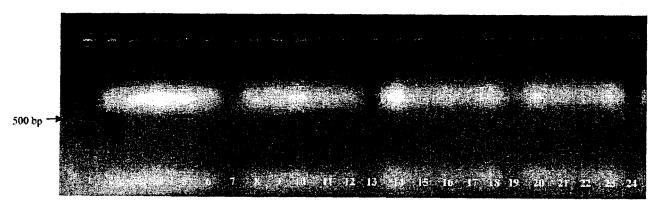


Fig. 3. RT-PCR 2 products from blood donor sera. Several amplicons generated by RT-PCR 2 were of the expected size (lanes 5, 10, 19, and 22). Lanes 1, 13, and 24: molecular weight ladder. Lane 2: positive-control RT-PCR 2. Lanes 3, 11, and 20 are negative controls containing all RT-PCR reagents without test specimen.

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The sixth polymerase region amplicon (N104), was closest (68% identity) to SMSV-13 and 44% identical to F9. The capsid gene sequence from RT-PCR 1 best (97% identity) matched SMSV-5.

A smaller nucleotide (113 nt) region of the six polymerase region amplicons was shared with 22 SMSV-like Vesivirus entries in GenBank (Fig. 4). The five similar polymerase region amplicons were distinct from each of the GenBank entries and diverged from published Vesivirus sequences for the most part at sites where other Vesivirus strains also differed from each other (Part A). The amplicon from donor N104 also had a Vesivirus sequence, but was distinct from other Vesivirus strains, including F9, differing from published Vesivirus most at the 5' end of the sequence, in a manner akin to how FCV differed from SMSV-like Vesivirus strains. Strain Hom-1 derived from a skin vesicle of a previously reported case of human Vesivirus infection [Smith et al., 1998a] had a short (47-64 nt) region of overlap with the serum-derived polymerase region amplicons. In this region of overlap, nucleotide sequence identity with the polymerase region amplicons ranged from 66% to 83%.

DISCUSSION

In this report, the detection of antibodies to Vesivirus and of SMSV-like Vesivirus viremia in blood donors is described at a U.S. regional blood bank serving eight Northwestern states. A higher prevalence of anti-Vesivirus antibody was observed, in comparison with normal blood donors, in donors with elevated serum ALT values, and the highest prevalence, among the groups tested, in cases of clinical hepatitis of unknown etiology associated with transfusion or dialysis. In addition, Vesivirus RNA was detected in healthy and high ALT blood donors, occurring numerically but not statistically significantly more frequently in the high ALT blood donors than among the healthy donors tested. The findings of this study and the attributes of Vesivirus calicivirus strains in mammals extend the potential for Vesivirus disease in humans from a few well-described cases to a broader population.

The complementary evidence for viremia included positive results from separate laboratories testing separate sample sets, utilization of RT-PCR assays that amplified non-overlapping genomic regions, direct detection of genomic RNA by hybridization, and amplicon sequencing that revealed non-identity of sequence compared with known Vesivirus, including strains characterized in one of the laboratories previously [Rinehart-Kim et al., 1999] and among the set of characterized amplicons themselves. These results together indicate that the positive laboratory results were not because of contamination in sample collection and handling.

The antigens utilized for detection of anti-Vesivirus antibody were purified by cesium chloride banding, which yields a homogenous population of viable particles. A pool of three SMSV strains was utilized, with each strain representing a different potential mechanism for exposure of humans to *Vesivirus*: SMSV-5 detected previously in a human case, SMSV-13 known to cause disease in two livestock species, and SMSV-17 recovered from edible shellfish. The separate phylogenetic cluster within the *Vesivirus* genus defined by FCV was represented by a live vaccine strain widely administered to cats. The noted differences among serum groups in patterns of estimated prevalence remained across a broad range of potential assay cutpoints. It would be unexpected for humans to have *Vesivirus* viremia, but not exhibit serologic evidence of infection.

Viremia was expected to be part of the natural history of Vesivirus infections in humans because one of two cases described had a disseminated vesicular exanthem of the hands and feet from which Vesivirus was cultivated [Smith et al., 1998a]. The Vesivirus strains causing Vesivirus viremia in this study were closest by genome sequence comparison to SMSV, marine Vesivirus. SMSV Vesivirus are widely distributed in marine and some terrestrial animals and may routinely "traffic" among these hosts, cycling from their large marine reservoirs onto land and perhaps back again. An ocean presence has been established by the isolation of virus, the presence of specific neutralizing antibodies, or by genome amplification and sequencing for 43 of the 46 known serotypes of the genus Vesivirus, including serotypes isolated initially from terrestrial hosts and named feline (FCV), primate (PCV Pan-1), bovine (BCV Bos-1), reptilian (RCV Cro-1), swine (VESV-A48-K56), mink (MCV), and human (SMSV-5 Hom-1 and HuCV Hom-1) caliciviruses [Evermann et al., 1983; Smith et al., 1983, 1998a,b, 2002b; Seal et al., 1995; Reid et al., 1999]. Another marine Vesivirus isolated from walrus (WCV) causes hepatitis in domestic animals [Smith, 2000; Ganova-Raeva et al., 2004]. Vesivirus caliciviruses are resistant to environmental degradation; stable in aquatic substrates; multiply to high titer, with an estimated 10¹³ virions released into the ocean daily by a single California gray whale (Eschricitus gibbosus) [Smith et al., 1998b, 2004] and, in the case of FCV, have a cosmopolitan distribution [Studdert, 1978; Smith et al., 1998bl. Such attributes indicate the potential for frequent contact between a diversity of Vesivirus biotypes and hosts.

Known examples of such interaction include mussels and oysters, aquatic filter feeders that concentrate particulates, including viruses, from the water column, preserve viral viability for 60 days or more and thereby can deliver large doses of viable Vesivirus to species ingesting contaminated shellfish [Smith, 2000; Burkhardt et al., 2002]. Another example is from the mid-Pacific (French Frigate Shoals), where Vesivirus-infected fingerlings of two fish genera (Aterinomorous spp. and Encrasicolina spp.) were eaten by white term (Gygis alba) hatchlings that developed a Vesivirus-associated blistering disease of the feet [Poet et al., 1996]. As mentioned above, whales (California gray) that can shed large numbers of Vesivirus particles per

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Vesivirus Strain	Nucleotide Sequence
chimpanzee Pan-1	ACCACTCATA TCATCTGTCA TGCCCAAAGT CTTCACCAAC CTGAAACAGT TTGGTCTGAA ACCGACCCGG ACCGACAAAA CGGATGCTGA GATAACGCCT ATCCCTGCTG ATG
Study N330	
Study N102	
Study 310	0 -T
Study 214	G
Study 298	
SMSV-6	GC
VESV E54	CAC
VESV A48	GGC
SMSV-4A	GC
skunk 4-2S	GCT
SMSV-4B	GC
VESV 1934b	C
skunk 4-1L	GC
skunk 7-2	GC
SMSV-1A	
SMSV-2	G
SMSV-1B	
SMSV-5	TCTTG-GACGCCTTAA-
walrus	TCAC-AAA
SMSV-7	TCC-AA
rabbit	
VESV C52	C
bovine	CCA-ACA-A-
VESV 155	CCCAC
SMSV-14	CACACAC
SMSV-13	TCT
Study N104	GCGAGGCT CAT-G-CGGT C-G-GG GGGGGAG
feline F9	TATTA-GTAT GAG-A-T- GTGA-CA- TTGGATTTCTTCC- AC GTTGTTGA-C ATGATGA-CCT

Fig. 4. Nucleotide (A) and amino acid (B) alignment of shared sequence among *Vesivirus* polymerase region amplicons from blood of study subjects and from *Vesivirus* strains represented in GenBank. Study strain sequences shared with these GenBank entries are aligned and placed next to the strains with which they had the highest sequence identity. GenBank sequences are grouped according to similarity to each other. Five of the study strains were similar to each other and closest to chimpanzee *Pan-1* and San Miguel sea lion serotype 6 strains. The other polymerase study strain also was closest to known *Vesivirus*, but in the pattern of sequence homology similar to feline calicivirus.

В	
Vesivirus Strain	Amino Acid Sequence
chimpanzee Pan-1	PLISSVMPKV FTNLKQFGLK PTRTDKTDAE ITPIPAD
Study N330	***************************************
Study N102	**********
Study 310	R
Study 214	A
Study 298	I
SMSV-6	
VESV E54	TA
VESV A48	
SMSV-4A	NA
skunk 4-2S	
SMSV-4B	
VESV 1934b	
skunk 4-1L	
skunk 7-2 & 3L	
SMSV-IA	
SMSV-2	
SMSV-1B	
SMSV-5	
walrus	к-
SMSV-7	к-
rabbit	
VESV C52	Т-
bovine	Т-
VESV 155	
SMSV-14	LA
SMSV-13	RL
Study N104	-EAHCRSAK-GEN
feline F9	IMYA-ISDOI -GSSYVSVGA -EDP-

Fig. 4. (Continued)

gram of feces per day also migrate thousands of miles annually between the Sea of Cortez and the Arctic Ocean [Akers et al., 1974; Smith et al., 2004]. Vesivirus has been recovered at titers of 10^7 infectious virions per gram of spleen in naturally and experimentally infected opal-eye fish (Girella nigricans). These fish are resident along the Southern California coast [Smith et al., 1980a,b, 1981] and are a sports and commercial fish and a common food source for seals, some of which also have extensive migration cycle. Seals reproduce on land or ice, where Vesivirus-induced reproductive failure and death occur, where foraging scavengers can further redistribute the viruses into terrestrial ecosystems, and where exposure to Vesivirus likely occurred for one human case [Smith et al., 1998a].

The present findings indicate a broader potential for Vesivirus infection and, perhaps, illness in humans than previously recognized. The strains causing viremia and to which antibody was detected in this study are similar to the Vesivirus with an ocean reservoir. A finding of subclinical viremia and the detected highest seroprevalence in cases of clinical hepatitis associated with transfusion or dialysis suggest that blood exposure that may have led to hepatitis also could lead to higher exposure to Vesivirus. If Vesivirus causes hepatitis in humans, then an estimated rate for such causation can

be derived from the study findings, as follows: the rate of 10 of 91 donors amplicon-positive in the serum who also had elevated serum liver transaminases (ALT) values, together with the rate of high ALT values occurring in about 1 in 1,000 blood donors, would correspond to a rate of \sim 1 in 10,000 blood donors who might have active, subclinical Vesivirus hepatitis. The association of higher anti-Vesivirus antibody prevalence with clinical hepatitis of unknown etiology would require further study. The finding of Vesivirus viremia in otherwise normal blood donors indicates that blood exposure to caliciviruses of the genus Vesivirus could occur by multiple routes of exposure. The diversity of host species, mechanisms of exposure and tissue tropisms for Vesivirus with the findings of this study suggest additional Vesivirus disease manifestations might be found in humans with further investigation.

ACKNOWLEDGMENTS

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REFERENCES

- Akers TG, Smith AW, Latham AB, Watkins HM. 1974. Calicivirus antibodies in California gray whales (Eschrichtius robustus) and Steller sea lions (Eumetopias jupatus). Arch Gesamte Virusforsch 46:175-177.
- Barlough JE, Berry ES, Skilling DE, Smith AW, Fay FH. 1986. Antibodies to marine caliciviruses in the Pacific walrus (Odobenus rosmarus divergens Illiger). J Wildl Dis 22:165-168.
- Barlough JE, Matson DO, Skilling DE, Berke T, Berry ES, Brown RF, Smith AW. 1998. Isolation of reptilian calicivirus Crotalus type 1 from feral pinnipeds. J Wildl Dis 34:451–456.
- Berke T, Golding B, Jiang X, Cubitt DW, Wolfaardt M, Smith AW, Matson DO. 1997. Phylogenetic analysis of the Caliciviruses. J Med Virol 52:419-424.
- Berry ES, Skilling DE, Barlough JE, Vedros NA, Gage LJ, Smith AW. 1990. New marine calicivirus serotype infective for swine. Am J Vet Res 51:1184-1187.
- Burkhardt W III, Blackstone GM, Skilling D, Smith AW. 2002. Applied technique for increasing calicivirus detection in shellfish extracts. J Appl Microbiol 93:235–240.
- Culley AI, Lang AS, Suttle CA. 2003. High diversity of unknown picorna-like viruses in the sea. Nature 424:1054-1057.
- Dean AG, Dean JA, Coulombier D, Brendel KA, Smith DC, Burton AH,
 Dicker RC, Sullivan K, Fagan RF, Arner TG. 1994. Epi Info, Version
 6: A word processing, database, and statistics program for
 epidemiology on microcomputers. Atlanta, Georgia, Centers for
 Disease Control and Prevention.
- Enserink M. 2000. Emerging diseases. Malaysian researchers trace Nipah virus outbreak to bats. Science 289:518-519.
- Enserink M. 2003. SARS in China. China's missed chance. Science 301:294-296.
- Evermann JF, Smith AW, Skilling DE, McKeirnan AJ. 1983. Ultrastructure of newly recognized caliciviruses of the dog and mink. Arch Virol 76:257-261.
- Ganova-Raeva L, Smith AW, Fields H, Khudyakov Y. 2004. New Calicivirus isolated from walrus. Virus Res 102:207-213.
- Green KY, Ando T, Balayan MS, Berke T, Clarke IN, Estes MK, Matson DO, Nakata S, Neill JD, Studdert MJ, Thiel HJ. 2000. Taxonomy of the caliciviruses. J Infect Dis 181:S322—S330.
- Kurth A, Everman JF, Skilling DE, Matson DO, Smith AW. 2006. Prevalence of vesivirus in a laboratory-based set of serum samples obtained from dairy and beef cattle. Am J Vet Res 67:114-119.
- Lang AS, Culley AI, Suttle CA. 2004. Genome sequence and characterization of a virus (HaRNAV) related to picorna-like viruses that infects the marine toxic bloom-forming alga Heterosigma akashiwo. Virology 320:206-217.
- Matson DO. 1999. Re-analysis of serologic data from the Australian study of human health risks of infection by rabbit hemorrhagic disease virus. Royal Society of New Zealand, Wellington: Misc.-Series 55:62-66.
- Matson DO, Berke T, Dinulos MB, Poet E, Zhong WM, Dai XM, Jiang X, Golding B, Smith AW. 1996. Partial characterization of the genome of nine animal caliciviruses. Arch Virol 141:2443–2456.
- Meanger J, Carter MJ, Gaskell RM, Turner PC. 1992. Cloning and sequence determination of the feline calicivirus strain F9. Biochem Soc Trans 20:26S-.
- Neill JD, Meyer RF, Seal BS. 1995. Genetic relatedness of the caliciviruses: San Miguel sea lion and vesicular exanthema of swine viruses constitute a single genotype within the Caliciviridae. J Virol 69:4484-4488.
- Neill JD, Meyer RF, Seal BS. 1998. The capsid protein of vesicular exanthema of swine virus serotype A48: Relationship to the

- capsid protein of other animal caliciviruses. Virus Res 54: 39-50.
- Poet SE, Skilling DE, Megyesl JL, Gilmartin WG, Smith AW. 1996. Detection of a non-cultivatable calicivirus from the white tern (Gygis alba rothschildi). J Wildl Dis 32:461-467.
- Reid SM, Ansell DM, Ferris NP, Hutchings GH, Knowles NJ, Smith AW. 1999. Development of a reverse transcription polymerase chain reaction procedure for the detection of marine caliciviruses with potential application for nucleotide sequencing. J Virol Methods 82:99-107.
- Rinehart-Kim JE, Zhong WM, Jiang X, Smith AW, Matson DO. 1999. Complete nucleotide sequence and genomic organization of a primate calicivirus, Pan-1. Arch Virol 144:199-208.
- Seal BS, Lutze-Wallace C, Kreutz LC, Sapp T, Dulac GC, Neill JD. 1995. Isolation of caliciviruses from skunks that are antigenically and genotypically related to San Miguel sea lion virus. Virus Res 37:1-12.
- Smith AW. 2000. Virus cycles in aquatic mammals, poikilotherms, and invertebrates. In: Hurst C, editor. Viral ecology. San Diego: Academic Press, pp 447-491.
- Smith AW, Boyt PM. 1990. Caliciviruses of ocean origin: A review. J Zoo Wildl Med 21:3-23.
- Smith AW, Prato CM, Skilling DE. 1977. Characterization of two new serotypes of San Miguel sea lion virus. Intervirology 8:30-36.
- Smith AW, Vedros NA, Akers TG, Gilmartin WG. 1978. Hazards of disease transfer from marine mammals to land mammals: Review and recent findings. J Am Vet Med Assoc 173:1131-1133.
- Smith AW, Skilling DE, Brown RJ. 1980a. Preliminary investigation of a possible lung worm (Parafilaroides decorus), fish (Girella nigricans), and marine mammal (Callorhinus ursinus) cycle for San Miguel sea lion virus type 5. Am J Vet Res 41:1846-1850.
- Smith AW, Skilling DE, Dardiri AH, Latham AB. 1980b. Calicivirus pathogenic for swine: A new serotype isolated from opaleye Girella nigricans, an ocean fish. Science 209:940-941.
- Smith AW, Skilling DE, Prato CM, Bray HL. 1981. Calcivirus (SMSV-5) infection in experimentally inoculated Opaleye fish (Girella nigricans). Arch Virol 67:165-168.
- Smith AW, Mattson DE, Skilling DE, Schmitz JA. 1983. Isolation and partial characterization of a calicivirus from calves. Am J Vet Res 44:851–855.
- Smith AW, Berry ES, Skilling DE, Barlough JE, Poet SE, Berke T, Mead J, Matson DO. 1998a. In vitro isolation and characterization of a calicivirus causing a vesicular disease of the hands and feet. Clin Infect Dis 26:434-439.
- Smith AW, Skilling DE, Cherry N, Mead JH, Matson DO. 1998b. Calicivirus emergence from ocean reservoirs: Zoonotic and interspecies movements. Emerg Infect Dis 4:13-20.
- Smith AW, Matson DO, Stein DA, Skilling DE, Kroeker AD, Berke T, Iversen PL. 2002a. Antisense treatment of caliciviridae: An emerging disease agent of animals and humans. Curr Opin Mol Ther 4:177-184.
- Smith AW, Skilling DE, Matson DO, Kroeker AD, Stein DA, Berke T, Iversen PL. 2002b. Detection of vesicular exanthema of swine-like calicivirus in tissues from a naturally infected spontaneously aborted bovine fetus. J Am Vet Med Assoc 220:455-458.
- Smith AW, Skilling DE, Castello JD, Rogers SO. 2004. Ice as a reservoir for pathogenic human viruses: Specifically, caliciviruses, influenza viruses, and enteroviruses. Med Hypotheses 63:560–566.
- Studdert MJ. 1978. Caliciviruses. Brief review. Arch Virol 58:157-191.
- Trampuz A, Prabhu RM, Smith TF, Baddour LM. 2004. Avian influenza: A new pandemic threat? Mayo Clin Proc 79:523– 530.

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	:	なし	該当	2006. 2. 15		·		識別番号・報告回数			
		公表国	040 2006			- 結人血漿	新鮮凍約	一般的名称			
			ProMED 20060205-0040, 2006 Feb 3. 情報源:[1]BBC News, UK, 2006 Feb 3. [2]Reuters alert. [3]Le Mauricien, 2006 Feb 3.		研究報告の公表状況			販売名(企業名)			
の注意記載状況・ 他参考事項等			○チクングンヤーレユニオンとモーリシャス [1]レユニオン島でチクングンヤウイルスが猛烈な勢いで広まり、患者数は2006年1月下旬の1週間だけで1万5千人増え計5万人								
	新鮮凍結血漿「日	// 重型////// 工即/ /									
等の感染	血液を介するウイ 細菌、原虫等の感 vCJD等の伝播の!	知 (Aedes albopictus)の駆除を決定した。1月から2月2日までに15例が報告されている。 細菌、原虫等									
	概 こしつる致死的疾患に限られる、と語った。保健者は全てのホケルと医師に疑わしい症例を報告するよう水のた。 モーリシャスでは2005年5月にチクングンヤの流行があり、週に数百名の患者が発生した。患者は6月には減り始め、9月までには散見される程度にまで減少した。2006年1月の患者の多くは最近レユニオンに行った人であった。ところが最近10日ほどは渡航歴のない患者が増えている。幾人かの血液検体からウイルスが検出された。Mahebourgでの集団発生を別にすると患者は様々な地域で発生している。										
	•	報告企業の意見 今後の対応							-		
		日本赤十字社では、輸血感染症対策として問診時に海外渡航歴の 有無を確認し、帰国後4週間は献血不適としている。今後も引き続き、 新たなウイルス等による感染症の発生状況等に関する情報の収集に 努める。									

ProMED情報(詳細)

1

記事番号	20060205-0040
重要度	C
タイトル	PROChikungunya – Mauritius and Reunion Island (07): Reunion
感染症名	チクングンヤ
主症状	
日付	2006/02/03
流行国	
和訳概要	Fクングンヤーモーリシャスとレユニオン(07):レユニオン# [11]レユニオン 情報源:BBC News レユニオン島でチクングンヤが猛烈な勢いで広まり、患者数は先週だけで1万5千人で計5万人に達した。この疾患は致死的ではないが高熱と激痛をもたらす。治療法はなく、ワクチンもない。軍隊が派遣され、近日中に蚊の大量駆除が全島で行われる予定である。[モデレータ注:1週間に15000例とは驚異的である。読者には地理と気候について興味があるだろう。レユニオンはマダガスカルの東に位置し、マダガスカル諸島の一部である。赤道と南回帰線との間にあるので海洋熱帯性気候である。季節は涼しい乾季(5-10月)と暖かい雨季(11-4月)の二つに分かれる。今は雨季の真中であり、蚊族が最も活動的で多いときである。1946年までフランス領であり、現在はフランスの海外県] [2]レユニオン情報源:Reuters alert 入院管理局は9歳の患者がチクングンヤで死亡した、と発表した。 [3]モーリシャス 情報源:Le Mauricien モーリシャス当局はMahebourgにおいてチクングンヤの新規患者2例を確認し、ウイルスを媒介するヒトスジシマカ(Aedes albopictus)を駆除することを決定した。1月から2月2日までに15例が報告されている。 政府はヒトに感染を起こしうる蚊族を全て駆除するために機器を購入している。たとえ保健検査局が港湾や空港で目を光らせても、レユニオンからの旅客を検疫する協定は存在しない。保健省報道官は人を検疫する方法はあるが、対象は島インフルエンザとヒトーヒ感染を起こしうる致死的疾患に限られる、と語った。保健省は全ホテルと全医師に疑わしい症例を報告するよう求めた。 Dr. MI Issack 病理学者(細菌学)の個人的コメントモーリシャスでは2005年5月にチクングンヤのアウトブレイクがあり、週に数百名の患者が発生した。患者は6月には減り始め、9月までには時折みられる程度にまでなった。2006年1月の患者の多くは最近レユニオンに行った人たちであった。ところが最近10日ほどは渡航屋のない患者が増えている。幾人かの血液検体からウイルスが検出された。Mahebourgでの集団発生を別にすると他の患者は様々な地域で発生している。これまでのところ、チクングンヤの状況はレユニオンに比べればはるかに良い。2005年12月は大変乾燥しており、夏の豪雨は1月23日まで始まらなかった。

情報詳細【和文】

チクングンヤーモーリシャスとレユニオン(07):レユニオン#

[1]レユニオン

情報源:BBC News

レユニオン島でチクングンヤが猛烈な勢いで広まり、患者数は先週だけで1万5千人で計5万人に達した。この疾患は致死的ではないが高熱と激痛をもたらす。治療法はなく、ワクチンもない。軍隊が派遣され、近日 -70-

1

中に蚊の大量駆除が全島で行われる予定である。

[モデレータ注:1週間に15000例とは驚異的である。読者には地理と気候について興味があるだろう。レユニオンはマダガスカルの東に位置し、マダガスカル諸島の一部である。赤道と南回帰線との間にあるので海洋熱帯性気候である。季節は涼しい乾季(5-10月)と暖かい雨季(11-4月)の二つに分かれる。今は雨季の真中であり、蚊族が最も活動的で多いときである。1946年までフランス領であり、現在はフランスの海外県] [2]レユニオン

情報源:Reuters alert

入院管理局は9歳の患者がチクングンヤで死亡した、と発表した。

[3]モーリシャス

情報源:Le Mauricien

モーリシャス当局はMahebourgにおいてチクングンヤの新規患者2例を確認し、ウイルスを媒介するヒトスジシマカ(Aedes albopictus)を駆除することを決定した。1月から2月2日までに15例が報告されている。政府はヒトに感染を起こしうる蚊族を全て駆除するために機器を購入している。たとえ保健検査局が港湾や空港で目を光らせても、レユニオンからの旅客を検疫する協定は存在しない。保健省報道官は人を検疫する方法はあるが、対象は鳥インフルエンザとヒトーヒト感染を起こしうる致死的疾患に限られる、と語った。保健省は全ホテルと全医師に疑わしい症例を報告するよう求めた。

Dr. MI Issack 病理学者(細菌学)の個人的コメント

モーリシャスでは2005年5月にチクングンヤのアウトブレイクがあり、週に数百名の患者が発生した。患者は6月には減り始め、9月までには時折みられる程度にまでなった。2006年1月の患者の多くは最近レユニオンに行った人たちであった。ところが最近10日ほどは渡航歴のない患者が増えている。幾人かの血液検体からウイルスが検出された。Mahebourgでの集団発生を別にすると他の患者は様々な地域で発生している。これまでのところ、チクングンヤの状況はレユニオンに比べればはるかに良い。2005年12月は大変乾燥しており、夏の豪雨は1月23日まで始まらなかった。

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Archive Number 20060204.0358
Published Date 04-FEB-2006

Subject PRO/EDR> Chikungunya - Mauritius and Reunion Island (07): Reunion

CHIKUNGUNYA - MAURITIUS AND REUNION ISLAND (07): REUNION

A ProMED-mail post

<http://www.promedmail.org>
ProMED-mail is a program of the

International Society for Infectious Diseases

<http://www.isid.org>

In this issue:

- [1] Reunion update
- [2] Reunion fatal case
- [3] Mauritius

[1] Reunion

Date: Fri, 3 Feb 2006 05:56:22 -0500 (EST)
From: A-Lan Banks <A-Lan.Banks@thomson.com>
Source: BBC News, UK, 3 Feb 2006 [edited]

http://news.bbc.co.uk/1/hi/world/europe/4674376.stm

A crippling mosquito-borne disease is spreading at an accelerating rate on the French Indian Ocean island of Reunion, health officials say.

They say the number of cases of the viral illness, known as chikungunya, had risen to 50 000, an increase of 15 000 in the past week alone

The disease is not fatal, but those affected suffer high fever and severe pain. There is no cure or vaccine.

Hundreds of troops have been deployed on the island to eradicate mosquitoes.

Officials said the troops would be spraying the whole island against mosquitoes in the coming days.

The latest outbreak was first noticed there in February 2005 - but has spread at an accelerating rate since December.

Meanwhile, neighboring territories are mobilizing to contain the disease.

On the Seychelles - where 2,000 cases have been reported in the past 4 weeks - the army has been mobilized to exterminate mosquitoes, Reuters news agency reports.

The authorities in Madagascar also fear the disease may have reached their island, AFP news agency says.

Chikungunya fever is named after a Swahili word meaning "that which bends up" - referring to the stooped posture of those afflicted.

ProMED-mail cpromed@promedmail.org>

[The spreading speed at 15 000 cases per week, or 2143 per day, is incredible. A bit of geography and climate may be of interest to our readers. Reunion is located east of Madagascar and is a part of the Madagascar archipelago. Situated between the equator and the tropic of Capricorn, Reunion's climate is of tropical type (southern

hemisphere) with oceanic influence. The island is home to one of the world's most active volcanoes. There are 2 seasons: cool or dry season (May till October) and warm or wet season (November till April). Currently it is in the middle of the warm season when mosquitoes are most active and proliferative. The island was ruled as a colony until 1946, when it was made a "departement" (=department), or administrative unit, of France. - Mod.RY]

[2] Reunion - fatal case

Date: 4 Feb 2006

From: ProMED-mail promed@promedmail.org>

Source: Reuters alert [edited]

http://www.alertnet.org/thenews/newsdesk/L04459988.htm

A crippling mosquito-borne disease has claimed its first life and infected more than 50 000 people on the volcanic French island of Reunion, spooking tourists in the region, local authorities said.

The "chikungunya" disease, which is extremely painful and causes high fever, was not previously thought to be lethal and there is no known cure or vaccine.

The Reunion Regional Agency for Hospitalisation said late on Friday the virus was the only explanation for the death of a 9-year-old in January [2006].

The "chikungunya" fever is named after a Swahili word meaning "that which bends up," referring to the stooped posture of those afflicted.

First recognised in epidemic form in East Africa in 1952, it also leaves immune systems weak, providing opportunities for other diseases to set in.

The disease, which has already travelled to the nearby Indian Ocean islands of Seychelles, Mauritius and Mayotte, has prompted tourists to cancel bookings.

Since 23 Jan [2006], the Reunion Committee on Tourism said at least 1500 tours had been cancelled and the costs incurred from cancellations over the past 2 weeks were equal to its annual advertising budget. It did not provide any figures.

On Thursday [2 Feb 2006], Seychelles said it had mobilised its army to control the virus which has infected 2000 of the idyllic archipelago's 80 000 people.

Reunion last week earmarked \$720 000 to fight the outbreak, drafting 400 extra troops to help fight the mosquitoes that have spread the disease for nearly a year.

In neighbouring Mauritius, authorities are screening people at the airport and port, spraying places near hotels and guest houses and warning the public to take precautions.

The Reunion authorities said they planned to set up a scientific committee to better understand the disease. They said they had registered some 25 deaths citing the virus as a possible cause.

Authorities say people should remove stagnant water, use mosquito repellents and bed nets and spray bedrooms at night.

ProMED-mail

cpromed@promedmail.org>

[Fatalities are not commonly seen with chikungunya. More information on these above mentioned cases would be appreciated. - Mod.MPP]

[3] Mauritius

Date: Fri, 3 Feb 2006 14:43:53 -0500 (EST) From: Mohammad Issack <missack@intnet.mu>

Source: Le Mauricien, 3 Feb 2006 [trans. and edited]

Mauritian authorities are on high alert after the identification of 2 confirmed cases of Chikungunya in Mahebourg. A ministerial committee is chaired by the Prime Minister, Navin Ramgoolam, to decide on measures to take to prevent infection by the _Aedes albopictus_ mosquito, which transmits this viral illness that is characterized mainly by fever, muscular pain and joint swellings. 15 cases have been registered from January up to 2 Feb 2006; the ministry of Health in 2005 detected 3500 cases that it prefers to call suspected cases of Chikungunya.

The government is buying equipment for disinfection in order to destroy all mosquitoes liable to infect people. [Even if] the port and the airport are watched closely by the Health Inspectorate Division, there is no protocol to quarantine travellers arriving from Reunion Island. A spokesman for the Ministry of Health said this morning [3 Feb] that provisions for keeping people in quarantine exist [only] in protocols on avian influenza and in cases of a potentially fatal illness that is transmissible from person to person. The Ministry of Health is requesting all hotels and all medical practitioners in private practice to notify any suspect case.

Personal comment:

In Mauritius, the peak of the Chikungunya outbreak occurred in May [2005] when several hundred cases were reported per week. The number of cases started to fall in June and by September, only occasional cases were reported. Most cases reported in January [2006] occurred in people who had a history of recent travel to Reunion Island. However, in the [last] 10 days, some cases in people without history of travel have been reported. Some have been confirmed by virus isolation in tissue culture from blood samples. With the exception of the cluster in Mahebourg, the other cases live in different regions of Mauritius.

Up to now, the situation with Chikungunya virus has been much less dramatic than in Reunion Island. However, this may change. December [2005] was very dry in Mauritius, and heavy summer rainfall did not occur until 23 Jan 2006.

Dr. M. I. Issack
Pathologist (Microbiology)
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Mauritius
<missack@intnet.mu>

[ProMED is grateful for the timely information and comment given by Dr. M. I. Issack from Mauritius. In the wake of the extraordinary epidemic of chikungunya on Reunion Island, spread of the epidemic to other islands seems inevitable. - Mod.RY]

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[see also:
Chikungunya - Madagascar: susp., RFI 20060202.0340
Chikungunya - Mauritius and Reunion Island (06): Reunion 20060203.0343
Chikungunya - Mauritius and Reunion Island (05): Reunion 20060131.0306
Chikungunya - Mauritius and Reunion Island (04): Reunion 20060127.0254
Chikungunya - Mauritius and Reunion Island (03): Reunion 20060124.0230
Chikungunya - Mauritius and Reunion Island (02): Reunion 20060121.0202
Chikungunya - Mauritius and Reunion Island: Reunion 20060102.0007
2005
Chikungunya - Mauritius and Reunion Island (04): Reunion 20051231.3716
Chikungunya - Mayotte, Reunion, Comoros 20050913.2707
Chikungunya - Indonesia (Tangerang) 20050717.2059
Chikungunya - Mauritius and Reunion Island (03) 20050624.1770
Deaths at sea - France (Reunion Island): RFI
                                              20050622.1759
Chikungunya - Mauritius and Reunion Island (2) 20050520.1384
Chikungunya - Mauritius and Reunion Island 20050519.1372
Chikungunya - Indonesia (West Lombok)
                                              20050422.1121
Chikungunya - Comoros (Ngazidja)
                                              20050405.0986
Chikungunya - Sri Lanka (02)
                                              20050223.0581
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医薬品 医薬部外品

研究報告 調査報告書

化粧品

識別番号・報告回数			報告日 第一報入手 E 2006 年 2 月 28		新医	薬品等の区分 該当なし	国	
一般的名称 ①乾燥抗 HBs 人免疫グロブリン ②ポリエチレングリコール処理抗 HBs 人免疫		1ール処理抗 HBs 人免疫ク	プロブリン 研究報告の 公表状況	研究報告の ProMED2006022		公表国フランス		
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た。 フランス 研 ン島で死	保健省によると、チク :亡し、現在も人口の 2 0						ļ	
		報告企業の意	見			今後の対応	血漿を原料	
との報告であ 血漿分画製剤 ウイルスが指	る。 lからのチクングンヤウ	イルス伝播の事例は報告さ をモデルウイルスとしたウ	ンヤ感染が大量に発生し、列 れていない。また、万一原料 7イルスバリデーション試験原	A血漿にチクングンヤ	を与えな	本剤の安全性に いと考えるので、 とらない。		

使用上の注意記載状況・ その他参考事項等

表として静注用ヘブスプリン-IH の記載 示す。

- 重要な基本的注意
- 」)本剤の原材料となる血液については、HBs 抗原、抗 HCV 抗体、抗 HIV-1 抗体、抗 HIV-2 抗体陰性で、かつ ALT (GPT) 値 でスクリーニングを実施している。更に、 プールした試験血漿については、HIV-1、 HBV 及び HCV について核酸増幅検査 (NAT)を実施し、適合した血漿を本剤の製 造に使用しているが、当該 NAT の検出限 界以下のウイルスが混入している可能性 が常に存在する。本剤は、以上の検査に 適合した高力価の抗 HBs 抗体を含有する 血漿を原料として、Cohn の低温エタノー ル分画で得た画分からポリエチレングリ コール 4000 処理、DEAE セファデックス 処理等により抗 HBs 人免疫グロブリンを 濃縮・精製した製剤であり、ウイルス不 活化・除去を目的として、製造工程にお いて 60℃、10 時間の液状加熱処理及び濾 **過膜処理(ナノフィルトレーション)を** 施しているが、投与に際しては、次の点 に十分注意すること。



Archive Number 20060225.0619
Published Date 25-FEB-2006
Subject PRO/EDR> Chikungunya - Indian Ocean update (03): spread to France

A ProMED-mail post
http://www.promedmail.org
ProMED-mail is a program of the
International Society for Infectious Diseases
http://www.isid.org

Date: Sat, 25 Feb 2006 07:14:58 -0500 (EST)

From: Mary Marshall kropical.forestry@btinternet.com

Source: Reuters Alertnet, 25 Feb 2006 [edited]

http://www.alertnet.org/thenews/newsdesk/L25768324.htm

Doctors in mainland France have detected a mosquitoborne disease among people returning from the Indian Ocean region, where the virus is spreading rapidly, a senior health official said on Saturday.

France's health minister has blamed "Chikungunya" fever, for which there is no known cure or vaccine, for directly or indirectly killing 77 people on the French island of [the] Reunion off the south east coast of Africa. French health officials say 157 000 people have now been infected by the disease on Reunion, about one in 5 of the population.

"We have people returning from Reunion who have symptoms of chikungunya and their diagnoses have been confirmed," Francois Bricaire, head of the infectious diseases service at Pitie-Salpetriere hospital in Paris, told Europe 1 radio. "It's not surprising, quite simply because of the contacts between the island of Reunion and mainland France." He said about 30 cases had been found by his service and it was likely that other medical services had detected cases. The disease can only spread via mosquitoes and Bricaire did not say whether the people with symptoms were confined or allowed home.

Health minister Xavier Bertrand told Europe 1 that the mosquito which carries the virus could be present in south eastern France but gave no details. The illness, which has also been found in the nearby Indian Ocean islands of the Seychelles and Mauritius [and Mayotte. – Mod.RY], is marked by high fever and severe rashes. Most people recover but it is extremely painful.

The number of people infected in Mauritius has risen to 962 from 341 the previous week, the Mauritius government said.

French prime minister Dominique de Villepin is due to travel to Reunion on Sunday. He faces growing criticism over the failure to prevent the disease spreading and said this week that the entire island should be cleared of mosquitoes.