e.g. blood donors, raises a number of unresolved ethical questions. For instance, how should reactive test results be handled when obtained with the first available test, which cannot be confirmed or clarified by another method?

13. Exclusion of persons from donating blood

Donor selection criteria based on the history of the donor must still be used for risk prevention; an overview can be found in Appendix (E) of [1]. The regulations applicable in Germany have been adopted in Haemotherapy Guidelines [75].

In the past few years it has been discussed whether potential transmission by transfusion could lead to perpetuation of vCJD among humans, even though transmission through the food chain has been stopped, and whether an exclusion of transfusion recipients could essentially influence the course of the vCJD epidemic. In several European countries including the UK, the Netherlands, Switzerland and France—there albeit as early as 1998 under the impression of virus transmissions—the exclusion of transfusion recipients from donating has meanwhile been laid down.

In order to obtain a scientific database, a model calculation based on very pessimistic assumptions was prepared. The epidemiological model describes the spreading of an infection, in this case vCJD, due to blood donations based on the demographic situation in Germany. It assumes that 2,000 individuals were infected by contaminated food during a limited period of 10 years. The total population comprises 80 million people. The parameters for the model were estimated on the basis of four data sets:

- Donations from 262,071 donors at the blood donation services of DRK (DRK-Blutspendedienst) West Hagen, Germany:
- 2. 617 controls of a case control study on Creutzfeldt—Jakob Disease at Göttingen University, Germany;
- 3. age distribution of 1,343 transfusion recipients at the University Hospital of Essen, Germany; and
- 4. a longitudinal study from Newcastle on the survival of 2,888 patients after a blood transfusion in June 1994.

The age structured model uses 2-month intervals and takes into account the following conditions:

- The mandatory age limit for blood donors is between 18 and 68 years. Each blood donor undergoes an active phase of donor activity the duration of which depends on age.
- The risk of receiving a blood transfusion strongly depends on age and has its peak at approx. 70 years.
- Survival after a blood transfusion also strongly depends on age. The increased mortality rate of transfusion recipients reduces the risk of spreading by blood donations.
- The model takes into account the current mortality rates in the Federal Republic of Germany.
- A mean incubation period of 16 years with a standard deviation of 4 years was assumed for the infection.

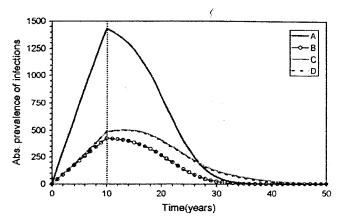


Fig. 3. Prevalence of vCJD. The vertical line shows the end of the phase of introduction of the infection by food. The curves show prevalence with the denominator of 80,000,000 German population, for the following cohorts: Curve A: Non-recipients (individuals without transfusion history, infection is therefore only possible by food). Curve B: Recipients (individuals with transfusion history), if transmission by transfusion is excluded (infection risk 0%, therefore—as in Curve A—infection possible only by food). Curve C: Recipients, if infected blood donations always lead to infection of recipients (infection risk 100%). The difference between Curve C and Curve B shows that the majority of infections was not caused by blood donations but by food. Curve D: as Curve C, but excluding donors with transfusion history. Reproduced from Fig. 2A in "How much would the exclusion of transfusion recipients from donating blood reduce the spread of vCJD?" Emerging Infectious Diseases, Vol. 13 No. 1, 2007.

The model permits exclusion of donors with a history of blood transfusions. It is assumed that 95% of the donors with a history of blood transfusions can be excluded.

Fig. 3 shows the absolute infection prevalence as a function of time predicted by the model. Prevalence increases during the 10-year period of food related infection and leads to a maximum of 1,434, infected individuals in the portion of the population without transfusion history (Curve A). In the portion of the population with transfusion history, depending on whether no risk of infection is assumed (0%, Curve B) or, in the most unfavorable case, an absolute infection risk is assumed (100%, Curve C) by blood donations from infected donors, 426 or 504 infected individuals, respectively, are to be expected. Maximum prevalence in the German population is 1,860 or 1,921 infected individuals respectively, corresponding to approx. 24 infected individuals per 1 million inhabitants. (The maximum value of 1,921 is slightly smaller than the total of 1,434 plus 504, since the maximum values of the individual curves are reached at different times.) The majority of infections caused by transfusions cannot be prevented by the exclusion of donors with a transfusion history, since they were infected by blood from food infected donors without a transfusion history. Thus, an exclusion of transfusion recipients would bring about only a minor contribution to prevention (Curves C and D can hardly be distinguished).

Given the initial rate of introduction of the infection, no further spreading occurs after that period of time, and, due to decreased life expectancies of vCJD infected individuals, the prevalence during the subsequent 20–30 years has a tendency

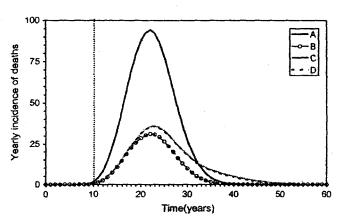


Fig. 4. Mortality from vCJD. The vertical line shows the end of the phase of introduction of the infection by food. The curves show incidence of deaths (Fig. 4) with the denominator of 80,000,000 German population, for the same cohorts A-D shown in Fig. 3. Reproduced from Fig. 3A in "How much would the exclusion of transfusion recipients from donating blood reduce the spread of vCJD?" *Emerging Infectious Diseases*, Vol. 13 No. 1, 2007.

towards zero again. Even if it is assumed that infected blood donations will always lead to infection of the recipient (infection risk = 100%), no further spreading occurs. In addition, the decrease in prevalence is only delayed due to the incubation period of the individuals infected (Curve C). An exclusion of transfusion recipients, even in the latter most pessimistic scenario, can bring about only a minor contribution to prevention (Curve D). This is also shown in Fig. 4, in which the annual incidence of deaths due to infection is shown. The maximum number of vCJD associated deaths occurs not before 23 years after the beginning of the onset of infection due to the long incubation period.

Fig. 5 compares the incidence of deaths of individuals infected by food (Curve E) with those that may be caused by blood donations with maximum risk of infection (Curve F). Due to the incubation period, transfusion associated deaths

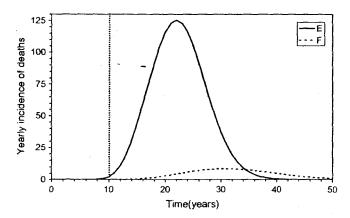


Fig. 5. Curve E: Deaths due to vCJD infections that could only be caused by food (totals of Curves A and B in Fig. 2). Curve F: Deaths due to vCJD caused by blood from infected donors (infection risk 100%) (difference between Curves C and B in Fig. 2). The vertical line marks the end of the phase of infection by food. Reproduced from Fig. 4A in "How much would the exclusion of transfusion recipients from donating blood reduce the spread of vCJD?" Emerging Infectious Diseases, Vol. 13 No. 1, 2007.

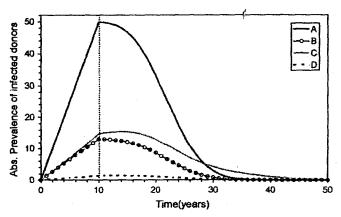


Fig. 6. Curve A: Prevalence of infected donors without transfusion history (infection possible only by food). Curve B: Prevalence of infected donors with transfusion history, if transmission by transfusion could be ruled out (infection risk 0%, therefore—as in A—infection possible only by food). Curve C: Prevalence of infected donors with transfusion history, if it is assumed that infected blood donations always lead to infection (infection risk 100%). The difference between Curve C and Curve B show that the majority of infections are not caused by blood donation but by food. Curve D: As Curve C, but excluding donors with transfusion history. This curve shows 5% of the donors with transfusion history, who were not excluded according to the model assumptions. The vertical line shows the end of the phase of infection by food. Reproduced from Fig. 5A in "How much would the exclusion of transfusion recipients from donating blood reduce the spread of vCJD?" Emerging Infectious Diseases, Vol. 13 No. 1, 2007.

occur markedly later than deaths caused by food infections. Within the displayed 50 years, 172 transfusion associated deaths have to be expected. During this period, however, a maximum of 15 cases could be prevented if donors with a transfusion history were excluded, equivalent to 1 case in 3-4 years. Out of the 2,000 individuals infected by food, we expect 1,557 vCJD cases if the infection risk of infected donors is 0%, and 1,729 cases if the infection risk is 100%. If approx. 20% of the donors were excluded, less than 1% of the cases would be prevented. Fig. 6 explains why the exclusion of donors with a transfusion history only slightly influences the incidence of deaths:

- The majority of the infected donors were infected by food and reveal no transfusion history (Curve A). This group is not covered by the exclusion criterion "donors with a transfusion history" and is able to continue to transmit the disease.
- Infected donors with a transfusion history can be excluded, but represent only a minor portion of infected donors (Curve B or C, respectively). The portion of donors infected by transfusions is very small (difference between Curve C and Curve B).

The assumptions chosen here present a considerable overestimation of the real risk of infection in Germany. Since an infection introduced by food cannot be sustained in the population, there is no further long-term risk after this route of transmission has been interrupted. Because of the low prevalence of approx. 24 infected individuals per 1 million (see above), linear reduction can be performed on predicted developments if markedly lower prevalences are assumed. The actual prevalence of individuals infected by food in Germany is probably lower by at least a factor of 10. Therefore, the above mentioned figures can probably be reduced by the corresponding factor. A detailed description of the model with all parameters and figures for the data sources on which this calculation is based has been published [105].

14. Impact of deferrals on the blood supply

Before introducing any donor deferrals, their effects should be quantified and a risk/benefit assessment (safety vs. blood supply) performed. It can be added to the discussion of the above model that an exclusion of transfusion recipients would not have prevented the three probable transmissions in the UK, since those donors were not transfusion recipients themselves. Moreover, it can be expected that an undetermined number of donors would not remember transfusions and continue to donate blood. On the other hand, as the French example shows, an exclusion of transfusion recipients is possible in principle, but would entail serious problems and disadvantages, would require major efforts and could therefore only be implemented over a longer period of time.

Examples from Germany may illustrate the impact of new exclusion criteria in general. With the 2000 update of guidelines, a rise of Hb limit for men from 125 to 135 g/l was introduced, which had a serious effect on the donor population. The deferral rate (Table 5) initially rose by approx. 2.5%, and after the male donors with Hb between 125 and 135 g/l had been deferred, decreased again to its initial level.9 Deferral rates tended to be rising from 322,312 in 2002 to 345,906 in 2003 (8.25% vs. 8.87% of the population prepared to donate). Since winter 2000/2001—despite intensive encouragement to donate blood-the demand for blood components, especially RBC, has not always been met so that, e.g., planned operations have to be postponed. Also, stricter donor exclusion criteria for the sake of improved safety may lead to a loss in donors, as anti-HBc testing [106] has led to permanent exclusion of approx. 0.6-1% of blood donors. New reasons for exclusion frequently are not understood by those concerned and require great educational efforts. As experience with the deferral due to a cumulative stay in the UK of more than 6 months between 1980 and 1996 has shown, considerable uncertainties remain despite numerous dialogues due to the lack of possibility to explore the donor's individual risk or to obtain a confirmatory/exclusion test. Exclusion of transfusion recipients could even be perceived as a signal that, despite all efforts, blood supplies are not sufficiently safe.

To secure blood supplies, new donors would have to be recruited to a considerable extent. According to the German data on epidemiology collected by the Robert Koch-Institut, the prevalence of virus infections in new blood donors is higher

Main reasons for deferral according to an analysis by the Institut für Transfusionsmedizin (Institute for Transfusion Medicine) Münster of the German Red Cross blood donation service West in 2004 (in % of volunteer donors)

2.21%
1.30%
1.17%
1.24%
0.78%
0.73%
0.72%
0.28%

than in long-term donors. While special promotion campaigns (cf. bone marrow/stem cell donors for children with leukemia) motivate many people to donate in the short-term, it is the continued reliability of donor preparedness in connection with the constantly required readiness to act that is important in blood donor promotion campaigns. The following possible approaches could secure continued donor preparedness and the supply of blood components:

- Economical use of blood and blood products: All measures that result in optimal use of blood and blood products will not only minimize the possible transmission risk but will also contribute to safeguarding the supply of blood and blood products. The activities in Germany and the European Union have been described above.
- Sustained recruitment measures: The GRC blood donation services are currently spending approx. €20 million a year on maintaining their existing donor base. Additional recruitment campaigns (approx. €3 million annually) have been aimed at the approx. 1-2% donor increase required to compensate for the annual rise in demand due to the increasing average age of the population. The GRC has been able to fulfill this goal with a relatively low budget, since advertising space in various media has generously been made available free of charge. In the case of donor exclusion due to transfusion history, just below 18,000 additional donors per month would be needed in the first half-year, and more than 11,800 additional donors per month in the second. Altogether, approx. 4 first-time volunteers would have to be recruited for each deferred donor (compare Appendix (E) of [1]). Such an additional recruitment campaign requiring more financial means could no longer be carried out by the blood donation services.
- Increasing the social prestige of the blood donors: It might be helpful to provide donors with a non-material reward in the form of increasing the social prestige attached to the act of giving blood. A professional study should explore the possibilities and develop suggestions for raising the esteem in which blood donors are held.

According to statistics from the German Red Cross blood donation services, the deferral rate among all volunteer donors in 2003 was 8.87%. There is a strong fluctuation in the deferral rate between the various blood donation services, which ranged

 $^{^{\}rm 9}$ German Red Cross (GRC) blood donation centre West, figures from North Rhine-Westphalia.

from 5-14% and is now 7-12%. No correlation can be detected between the deferral rate and the degree of conurbation. These differences can certainly not solely be explained by differences in the donor population of the German Red Cross blood donation services. An improvement could be the introduction of uniform interpretation aids throughout Germany.

15. Summary and conclusions

So far 162 cases of vCJD in the UK, 20 in France, 4 in Ireland, 2 in the Netherlands, 2 in the USA, and one case each in Canada, Italy, Japan, Portugal, Saudi Arabia, and Spain have been observed. Twenty-seven of these patients resided outside the UK and had not spent considerable time in the UK; a connection with stay in the UK is questionable in the case in Japan. It must not be ruled out that vCJD will be diagnosed in more countries. Recent model calculations in the UK [50] have resulted in lower than previously published estimates of the overall number of clinical vCJD cases, however, with considerable confidence intervals. A fundamental assumption is that new infections from the food chain have been effectively stopped. On the basis of new estimates, the number of up to 600 cases of vCJD for Germany indicated in the report of this group in 2001 can be considered as too pessimistic.

Three cases published in the UK since 2004 must be regarded as evidence for the transmissibility of the vCJD pathogen by blood. One of the recipients died from an unrelated disease. However, autopsy revealed the vCJD pathogen in the spleen and lymph nodes, pointing to a subclinical or not yet symptomatic infection. Unlike all other previously observed vCJD cases, this patient was heterozygous M/V at codon 129. This observation and the results from a serial investigation of appendix tissue in the UK could indicate that there are a considerable number of infected persons who might not develop vCJD, or in whom its manifestation is delayed. At present, it is not possible to ascertain whether infectivity is present in the blood of these persons, and if so, at what time and to what extent. Precautionary measures should therefore not be based exclusively on the number of already manifested vCJD cases and the forecast of future numbers of cases derived thereof.

A possible transmission of the vCJD pathogen by plasma products still cannot be entirely ruled out, but it seems unlikely since various experimental systems have shown that prions are largely removed during the manufacture of these blood products. Examining the effectiveness of these steps, however, should be continued in a product-oriented manner. A Note for Guidance was published in 2004 by the European Medicines Agency (EMEA) for this purpose [88].

Transmissibility of vCJD by transfusion had been assumed already in 2001; precautionary measures for minimizing the risk had been taken. As an additional measure, the exclusion of transfusion recipients from donating blood has to be considered in order to break a hypothetical chain of further spread and possible perpetuation of vCJD by blood products. Such exclusion has been introduced in the UK, the Netherlands, Switzerland, and as early as 1998—to prevent viral transmissions—in

France. A model calculation took these considerations into account; to be on the safe side, still a worst case scenario was dealt with. For Germany, the group concluded that such exclusion is not warranted. The evaluation of the potential gain in safety regarding vCJD, based on the above mentioned model calculation, was not considered to outweigh the drawbacks of such a measure.

The secondary route of infection by blood could largely be stopped as soon as a suitable test could be introduced into routine donor screening. No such test is currently available; developing and optimizing test methods should have high priority.

Acknowledgements

The excellent secretarial support by Mrs. Helene Knöß is gratefully acknowledged.

References

- [1] Report of the Working Group Commissioned by the German Federal Ministry of Health. Overall Blood Supply Strategy with Regard to Variant Creutzfeldt--Jakob Disease. Transfus Med Hemother 2006; 33(Suppl. 2).
- [2] Wells GA, Scott AC, Johnson CT, Gunning RF, Hancock RD, Jeffrey M, et al. A novel progressive spongiform encephalopathy in cattle. Vet Rec 1987;121(18):419-20.
- [3] Wilesmith JW, Ryan JB, Atkinson MJ. Bovine spongiform encephalopathy: epidemiological studies on the origin. Vet Rec 1991;128(9): 199-203.
- [4] Anderson RM, Donnelly CA, Ferguson NM, Woolhouse ME, Watt CJ, Udy HJ, et al. Transmission dynamics and epidemiology of BSE in British cattle. Nature 1996;382(6594):779-88.
- [5] Biacabe AG, Laplanche JL, Ryder L, Baron T. Distinct molecular phenotypes in bovine prion diseases. EMBO reports 2004;5:110-5.
- [6] Casalone C, Zanusso G, Acutis P, Ferrari S, Capucci L, Tagliavini F, et al. Identification of a second bovine amyloidotic spongiform encephalopathy: Molecular similarities with sporadic Creutzfeldt-Jakob disease. Proc Natl Acad Sci USA 2004;101:3065-70.
- [7] De Bosschere H, Roels S, Vanopdenbosch E. Atypical case of bovine spongiform encephalopathy in an East-Flemish cow in Belgium. J Appl Res Vet Med 2004;2:1-3.
- [8] Fraser H, Bruce ME, Chree A, McConnell I, Wells GA. Transmission of bovine spongiform encephalopathy and scrapie to mice. J Gen Virol 1992; 73(8):1891-7.
- [9] Wells GA, Hawkins SA, Green RB, Austin AR, Dexter I, Spencer YI, et al. Preliminary observations on the pathogenesis of experimental bovine spongiform encephalopathy (BSE): an update. Vet Rec 1998; 142(5):103-6.
- [10] Buschmann A, Groschup MH. Highly bovine spongiform encephalopathy-sensitive transgenic mice confirm the essential restriction of infectivity to the nervous system in clinically diseased cattle. J Infect Dis 2005;192:934-42.
- [11] Bruce ME. Strain typing studies of scrapie and BSE. In: Baker HF, Ridley RM, editors. Methods in molecular medicine: prion disease. Totowa, NJ: Humana Press; 1996. p. 223-36.
- [12] Bruce ME, Boyle A, Cousens S, McConnell I, Foster J, Goldmann W, et al. Strain characterization of natural sheep scrapie and comparison with BSE. J Gen Virol 2002;83:695-704.
- [13] Stack MJ, Chaplin MJ, Clark J. Differentiation of prion protein glycoforms from naturally occurring sheep scrapie, sheep-passaged scrapie strains (CH1641 and SSBP1), bovine spongiform encephalopathy (BSE) cases and Romney and Cheviot breed sheep experimentally

- inoculated with BSE using two monoclonal antibodies. Acta Neuropathol (Berl) 2002;104:279-86.
- [14] Lezmi S, Martin S, Simon S, Comoy E, Bencsik A, Deslys JP, et al. Comparative molecular analysis of the abnormal prion protein in field scrapic cases and experimental bovine spongiform encephalopathy in sheep by use of Western blotting and immunohistochemical methods. J Virol 2004;78:3654-62.
- [15] Thuring CM, Erkens JH, Jacobs JG, Bossers A, van Keulen LJ, Garssen GJ, et al. Discrimination between scrapie and bovine spongiform encephalopathy in sheep by molecular size, immunoreactivity, and glycoprofile of prion protein. J Clin Microbiol 2004;42:972-80.
- [16] Nonno R, Esposito E, Vaccari G, Conte M, Marcon S, Di Bari M, et al. Molecular analysis of cases of Italian sheep scrapie and comparison with cases of bovine spongiform encephalopathy (BSE) and experimental BSE in sheep. J Clin Microbiol 2003;41:4127-33.
- [17] Gretzschel A, Buschmann A, Eiden M, Ziegler U, Lühken G, Erhardt G, et al. Strain typing of German transmissible spongiform encephalopathies field cases in small ruminants by biochemical methods. J Vet Med B Infect Dis Vet Public Health 2005;52(2):55-63.
- [18] Benestad SL, Sarradin P, Thu B, Schonheit J, Tranulis MA, Bratberg B. Cases of scrapie with unusual features in Norway and designation of a new type, Nor98. Vet Rec 2003;153:202-8.
- [19] Bradley R. Experimental transmission of bovine spongiform encephalopathy. In: Court L, Dodet B, editors. Transmissible subacute spongiform encephalopathies: prion diseases. Paris: Elsevier; 1996. p. 51-6.
- [20] Andreoletti O, Berthon P, Marc D, Sarradin P, Grosclaude J, van Keulen L, et al. Early accumulation of PrP^(Sc) in gut-associated lymphoid and nervous tissues of susceptible sheep from a Romanov flock with natural scrapie. J Gen Virol 2000;81(Pt 12):3115-26.
- [21] Andreoletti O, Berthon P, Levavasseur E, Marc D, Lantier F, Monks E, et al. Phenotyping of protein-prion (PrPSc)-accumulating cells in lymphoid and neural tissues of naturally scrapic-affected sheep by double-labeling immunohistochemistry. J Histochem Cytochem 2002; 50(10):1357-70.
- [22] Schreuder BE, van Keulen LJ, Vromans ME, Langeveld JP, Smits MA. Tonsillar biopsy and PrPSc detection in the preclinical diagnosis of scrapie. Vet Rec 1998;142(21):564-8.
- [23] Will RG, Ironside JW, Zeidler M, Cousens SN, Estibeiro K, Alpérovitch A, et al. A new variant of Creutzfeldt-Jakob disease in the UK. Lancet 1996;347:921-5.
- [24] Will RG, Knight RS, Zeidler M, Stewart G, Ironside JW, Cousens SN, et al. Reporting of suspect new variant Creutzfeldt—Jakob disease. Lancet 1997;349:847.
- [25] Collinge J. Variant Creutzfeldt-Jakob disease. Lancet 1999;354: 317-23.
- [26] Collinge J, Sidle KC, Meads J, Ironside J, Hill AF. Molecular analysis of prion strain variation and the aetiology of 'new variant' CJD. Nature 1996;383:685-90.
- [27] Hill AF, Desbruslais M, Joiner S, Sidle KC, Gowland I, Collinge J, et al. The same prion strain causes vCJD and BSE. Nature 1997;389: 448-50, 526.
- [28] Bruce ME, Will RG, Ironside JW, McConnell I, Drummond D, Suttie A, et al. Transmissions to mice indicate that the 'new variant' CJD is caused by BSE agent. Nature 1997;389:498-501.
- [29] Lasmézas CI, Fournier JG, Nouvel V, Boe H, Marce D, Lamoury F, et al. Adaptation of the bovine spongiform encephalopathy agents to primates and comparison with Creutzfeldt-Jakob disease: implications for human health. Proc Natl Acad Sci USA 2001;98:4142-7.
- [30] Lasmézas CI, Deslys JP, Demaimay R, Adjou KT, Lamoury F, Dormont D, et al. BSE transmission to macaques. Nature 1996;381: 743-4.
- [31] Scott MR, Will R, Ironside J, Nguyen HO, Tremblay P, DeArmond SJ, et al. Compelling transgenic evidence for transmission of bovine spongiform encephalopathy prions to humans. Proc Natl Acad Sci USA 1999;96:15137-42.
- [32] Ironside JW, McCardle L, Horsburgh A, Lim Z, Head MW. Pathological diagnosis of variant Creutzfeldt—Jakob disease. APMIS 2002:110:70...87

- [33] McBride PA, Schulz-Schaeffer WJ, Donaldson M, Bruce M, Diringer H, Kretzschmar HA, et al. Early spread of scrapie from the gastrointestinal tract to the central nervous system involves autonomic fibers of the splanchnic and vagus nerves. J Virol 2001;75(19): 9320-7.
- [34] Will RG. Acquired prion disease: iatrogenic CJD, variant CJD, kuru. Br Med Bull 2003;66:255-65.
- [35] Hill AF, Butterworth RJ, Joiner S, Jackson G, Rossor MN, Thomas DJ, et al. Investigation of variant Creutzfeldt-Jakob disease and other human prion diseases with tonsil biopsy samples. Lancet 1999;353: 183-9.
- [36] Task Force vCJK. Abschlussbericht: Die Variante der Creutzfeldt-Jakob-Krankheit (vCJK). Epidemiologie, Erkennung, Diagnostik und Prävention unter besonderer Berücksichtigung der Risikominimierung einer iatrogenen Übertragung durch Medizinprodukte, insbesondere chirurgische Instrumente. Bundesgesundheitsbl Gesundheitsforsch Gesundheitsschutz 2002;45:376-94.
- [37] Bertram J, Mielke M, Beekes M, Lemmer K, Baier M, Pauli G. Inaktivierung und Entfernung von Prionen bei der Aufbereitung von Medizin-produkten—Ein Beitrag zur Prüfung und Deklaration geeigneter Verfahren. Bundesgesundheitsbl Gesundheitsforsch Gesundheitsschutz 2004:47:36-40
- [38] Lemmer K, Mielke M, Pauli G, Beekes M. Decontamination of surgical instruments from prion proteins: in vitro studies on the detachment, destabilization and degradation of PrPSc bound to steel surfaces. J Gen Virol 2004;85(Pt 12):3805-16.
- [39] Diringer H. Creutzfeldt-Jakob disease. Lancet 1996;347:1332-3.
- [40] Alpérovitch A, Zerr I, Pocchiari M, Mitrova E, de Pedro Cuesta J, Hegyi I, et al. Codon 129 prion protein genotype and sporadic Creutzfeldt-Jakob disease. Lancet 1999;353:1673-4.
- [41] Kretzschmar HA. Die Pathologie und Genetik der Prionkrankheiten beim Menschen. In: Hörnlimann B, Riesner D, Kretzschmar H, editors. Prionen und Prionkrankheiten. Berlin/New York: de Gruyter; 2001. p. 207-24.
- [42] Ironside JW, Head MW. Variant Creutzfeldt—Jakob disease: risk of transmission by blood and blood products. Haemophilia 2004; 10(Suppl. 4):64—9.
- [43] Brandel JP, Preece M, Brown P, Croes E, Laplanche JL, Agid Y, et al. Distribution of codon 129 genotype in human growth hormone-treated CJD patients in France and the UK. Lancet 2003;362:128-30.
- [44] Ghani AC, Ferguson NM, Donnelly CA, Hagenaars TJ, Anderson RM. Estimation of the number of people incubating variant CJD. Lancet 1998;352:1353-4.
- [45] Ghani AC, Donnelly CA, Ferguson NM, Anderson RM. Predicted vCJD mortality in Great Britain. Nature 2000;406(6796):583-4.
- [46] Ghani AC, Donnelly CA, Ferguson NM, Anderson RM. Updated projections of future vCJD deaths in the UK. BMC Infectious Diseases 2003;3:4.
- [47] Hilton DA, Ghani AC, Conyers L, Edwards P, McCardle L, Penney M, et al. Accumulation of prion protein in tonsil and appendix: review of tissue samples. BMJ 2002;325(7365):633-4.
- [48] Hilton DA, Ghani AC, Conyers L, Edwards P, McCardle L, Ritchie D, et al. Prevalence of lymphoreticular prion protein accumulation in UK tissue samples. J Pathol 2004;203(3):733-9.
- [49] Ironside JW, Bishop MT, Connolly K, Hegazy D, Lowrie S, Le Grice M, et al. Variant Creutzfeldt—Jakob Disease: a prion protein genotype analysis of positive appendix tissue samples from a retrospective prevalence study. BMJ 2006;332:1186—8.
- [50] Clarke P, Ghani AC. Projections of the future course of the primary vCJD epidemic in the UK: inclusion of subclinical infection and the possibility of wider genetic susceptibility. J R Soc Interface 2005;2(2):19-31.
- [51] Harney MS, Ghani AC, Donnelly CA, Walsh RM, Walsh M, Howley R, et al. vCJD risk in the Republic of Ireland. BMC Infect Dis 2003;3:28.
- [52] Chadeau-Hyam M, Alpérovitch A. Risk of variant Creutzfeldt—Jakob disease in France. Int J Epidem 2005;34:46—52.
- [53] Agence Française de Sécurité Sanitaire des Produits de Santé (AFS-SAPS). Risk analysis of new variant Creutzfeldt—Jakob disease transmission by blood and blood products. Report 2000;1—21.

- [54] Brown P, Cervenakova L, Diringer H. Blood infectivity and the prospects for a diagnostic screening test in Creutzfeldt-Jakob disease. J Lab Clin Med 2001;137:5-13.
- [55] Löwer J. Die Risikoeinschätzung einer Übertragung von Prionkrankheiten durch Blut, Augenhornhaut und Hirnhaut. In: Hoernlimann B, Riesner D, Kretschmar H, editors. Prionen und Prionkrankheiten. Berlin/New York: Walter de Gruyter; 2001. pp 440-55.
- [56] Cervenakova L, Yakovleva O, McKenzie C, Kolchinsky S, McShane L, Drohan WN, et al. Similar levels of infectivity in the blood of mice infected with human-derived vCJD and GSS strains of transmissible spongiform encephalopathy. Transfusion 2003;43:1687-94.
- [57] Hunter N, Foster J, Chong A, McCutcheon S, Parnham D, Eaton S, et al. Transmission of prion diseases by blood transfusion. J Gen Virol 2002; 83:2897—905.
- [58] Ricketts MN, Brown P. Transmissible spongiform encephalopathy update and implications for blood safety. Clin Lab Med 2003;23: 129-37.
- [59] Llewelyn CA, Hewitt PE, Knight RSG, Amar K, Cousens S, Mackenzie J, et al. Possible transmission of variant Creutzfeldt—Jakob disease by blood transfusion. Lancet 2004;363:417—21.
- [60] Peden AH, Head MW, Ritchie DL, Bell JE, Ironside JW. Preclinical vCJD after blood transfusion in PRPN codon 129 heterozygous patient. Lancet 2004;364:527-9.
- [61] Wroe SJ, Pal S, Durrenajaf S, Harpreet H, Macfarlane R, Joiner S, et al. Clinical presentation and pre-mortem diagnosis of variant Creutzfeldt-Jakob disease associated with blood transfusion: a case report. Lancet 2006;368:2061-7.
- [62] Hewitt PE, Llewelyn CA, Mackenzie J, Will RG. Creutzfeldt-Jakob disease and blood transfusion: results of the UK Transfusion Medicine Epidemiological Review study. Vox Sang 2006;91:221-30.
- [63] Arbeitskreis Blut (National Advisory Committee 'Blood'). Procedures to be followed in cases of variant Creutzfeldt—Jakob Disease (vCJD) in connection with blood, plasma and blood products. (Recommendation 33). Bundesgesundheitsbl Gesundheitsforsch Gesundheitsschutz 2006; 49:396—8. http://www.rki.de/cln_011/nn_231708/EN/Content/Prevention/ NAC_Blood/V_33E.templateId=raw.property=publicationFile.pdf/V 33E.
- [64] European Commission. Directorate C7—Risk assessment (SCENIHR/003/05). Scientific Committee on Emerging and Newly Identified Health Risks (SCENIHR): Opinion on the Safety of human-derived products with regard to variant Creutzfeldt—Jakob Disease (Adopted by the SCENIHR during the 7th plenary meeting of 28—29 September 2005).
- [65] Collinge J, Whitfield J, McKintosh E, Beck J, Mead S, Thomas DJ, et al. Kuru in the 21st century—an acquired human prion disease with very long incubation periods. Lancet 2006;367:2068-74.
- [66] AFSSAPS, Analyse du risque de transmission de la variante de la maladie de Creutzfeldt—Jakob par produits de santé et par les tissus et fluides d'origine humaine, actualisation des données du rapport du groupe ad hoc de décembre 2000 rapport de février 2004 (http://agmed.sante. gouv.fr/htm/13/13000.htm).
- [67] Herzog C, Salès N, Etchgaray N, Charbonnier A, Freire S, Dormont D, et al. Tissue distribution of bovine spongiform encephalopathy agent in primates after intravenous or oral infection. Lancet 2004;363:422-8.
- [68] Gregori L, Maring JA, MacAuley C, Dunston B, Rentsch M, Kempf C, et al. Partitioning of TSE infectivity during ethanol fractionation of human plasma. Biologicals 2004;32:1-10.
- [69] Yakovleva O, Janiak A, McKenzie C, McShane L, Brown P, Cervenakowa L. Effect of protease treatment on plasma infectivity in variant Creutzfeld-Jakob disease mice. Transfusion 2004;44:1700-5.
- [70] DNV Consulting. Risk Assessment of vCJD Infectivity in Blood, http:// www.dnv.com/binaries/AppII_tcm4-74416.pdf; 2003.
- [71] Wadsworth JD, Joiner S, Hill AF, Campbell TA, Desbruslais M, Luthert PJ, et al. Tissue distribution of protease resistant prion protein in variant Creutzfeldt—Jakob disease using a highly sensitive immunoblotting assay. Lancet 2001;358:171-80.
- [72] Bruce ME, McConnell I, Will RG, Ironside JW. Detection of variant Creutzfeldt—Jakob disease infectivity in extraneural tissues. Lancet 2001: 358:208—9.

- [73] Brown P, Cervenakova L, McShane LM, Barber P, Rubenstein R, Drohan WN. Further studies of blood infectivity in an experimental model of transmissible spongiform encephalopathy, with an explanation of why blood components do not transmit Creutzfeldt-Jakob disease in humans. Transfusion 1999;39:1169-78.
- [74] Gregori L, McCombie N, Palmer D, Birch P, Sowemimo-Coker SO, Giulivi A, et al. Effectiveness of leucoreduction for removal of infectivity of transmissible spongiform encephalopathies from blood. Lancet 2004;364:529-31.
- [75] Bundesärztekammer und Paul-Ehrlich-Institut (German Medical Association and Paul-Ehrlich-Institut). Guidelines governing the collection of blood and blood components and the use of blood products (haemotherapy) pursuant to Sections 12 and 18 of the Transfusion act as amended in 2005 (in German). Federal Gazette 209a; 5 November 2005.
- [76] Stenland CJ, Lee DC, Brown P, Petteway SR, Rubenstein Jr R. Partitioning of human and sheep forms of the pathogenic prion protein during the purification of therapeutic proteins from human plasma. Transfusion 2002;42:1497-500.
- [77] Foster PR. Removal of TSE agents from blood products. Vox Sang 2004;87(Suppl. 2):7-10.
- [78] Lee DC, Stenland CJ, Miller JL, Cai K, Ford E, Gilligan KJ, et al. A direct relationship between partitioning of the pathogenic prion protein and transmissible spongiform encephalopathy infectivity during the purification of plasma proteins. Transfusion 2001;41:449-55.
- [79] Vey M, Baron H, Weimer T, Gröner A. Purity of spiking agent affects partitioning of prions in plasma protein purification. Biologicals 2002; 30:187-96.
- [80] Tateishi J, Kitamoto T, Mohri S, Satoh S, Sato T, Shepherd A, et al. Scrapie removal using Planova[®] virus removal filters. Biologicals 2001:29:17-25.
- [81] Cai K, Miller JL, Stenland CJ, Gilligan KJ, Hartwell RC, Terry JC, et al. Solvent-dependent precipitation of prion protein. Biochim Biophys Acta 2002:1579:28-35.
- [82] Van Holten RW, Autenrieth SM. Evaluation of depth filtration to remove prion challenge from an immune globulin preparation. Vox Sang 2003;85:20-4.
- [83] Lee DC, Stenland CJ, Hartwell RC, Ford EK, Cai K, Miller JL, et al. Monitoring plasma processing steps with a sensitive Western blot assay for the detection of the prion protein. J Virol Methods 2000;84(1):77-89.
- [84] Foster PR. Assessment of the potential of plasma fractionation processes to remove causative agents of transmissible spongiform encephalopathy. Transfus Med 1999;9(1):3-14.
- [85] Foster PR, Welch AG, McLean C, Griffin BD, Hardy JC, Bartley A, et al. Studies on the removal of abnormal prion protein by processes used in the manufacture of human plasma products. Vox Sang 2000;78:86-95.
- [86] Reichl HE, Forster PR, Welch AG, Li Q, MacGregor IR, Somerville RA, et al. Studies on the removal of a bovine spongiform encephalopathy-derived agent by processes used in the manufacture of human immunoglobulin. Vox Sang 2002;83:137-45.
- [87] Foster PR, Griffin BD, Bienek C, McIntosh RV, MacGregor IR, Somerville RA, et al. Distribution of a bovine spongiform encephalopathy-derived agent over ion-exchange chromatography used in the preparation of concentrates of fibrinogen and factor VIII. Vox Sang 2004;86:92-9.
- [88] EMEA/410/01 Rev. 3: Note for Guidance on Minimising the Risk of Transmitting Animal Spongiform Encephalopathy Agents via Human and Veterinary Medicinal Products, (Released for consultation June 2004), http://www.emea.eu.int/index/indexh1.htm.
- [89] EMEA/410/01 Rev. 2: Note for Guidance on Minimising the Risk of Transmitting Animal Spongiform Encephalopathy Agents via Human and Veterinary Medicinal Products, (Adopted by CPMP/CVMP October 2003), http://www.emea.eu.int/index/indexh1.htm.
- [90] Frey L, Messmer K. Blood replacement in elective surgery: results of the Sanguis Study. Infusionsther Transfusionsmed 1993;20(Suppl. 2): 12-5.
- [91] The Sanguis Study Group. Use of blood products for elective surgery in 43 European Hospitals. Transfus Med 1994;4:251-68.

- [92] Capraro L, Nuutinen L, Myllyla G. Transfusion thresholds in common elective surgical procedures in Finland. Vox Sang 2000;78:96-100.
- [93] Morrison JC, Sumrall DD, Chevalier SP, Robinson SV, Morrison FS, Wiser WL. The effect of provider education on blood utilization practices. Am J Obstet Gynecol 1993;169:1240-5.
- [94] Hebért PC, Wells G, Blajchman MA, Marshall J, Martin C, Pagliarello G, et al. A multicenter, randomized, controlled clinical trial of transfusion requirements in critical care. Transfusion Requirements in Critical Care Investigators, Canadian Critical Care Trials Group. N Engl J Med 1999;340:409-17.
- [95] Arbeitskreis Blut (National Advisory Committee 'Blood'). Current recommendations for autologous haemotherapy (Recommendation 32). Bundesgesundheitsbl Gesundheitsforsch Gesundheitsschutz 2005;48: 700-2. http://www.rki.de/cln_011/nn_231708/EN/Content/Prevention/NAC_Blood/V_32E.templateId=raw.property=publicationFile.pdf/V_32E.
- [96] Blood Safety in the European Community: an initiative for optimal use. Conference Proceedings, W. Schramm, editor. European Commission, Strasbourg; 2000.
- [97] Bundesärztekammer (German Medical Association). Guidelines for therapy with blood components and plasma derivatives. 2nd revised ed. Köln: Deutscher Ärzteverlag, 3rd ed.; 2005
- [98] Council of Europe. Guide to the preparation, use and quality assurance of blood components (Recommendation No. R (95) 15) Council of Europe Publishing, Strasbourg (updated every year).
- [99] Soto C. Diagnosing prior diseases: needs, challenges and hopes. Nat Rev Microbiol 2004;2:809-19.

- [100] Safar JG, Geschwind MD, Deering É, Didorenko S, Sattavat M, Sanchez H, et al. Diagnosis of human prion disease. Proc Natl Acad Sci USA 2005;102:3501-6.
- [101] Saborio GP, Permanne B, Soto C. Sensitive detection of pathological prion protein by cyclic amplification of protein misfolding. Nature 2001; 411:810-3.
- [102] Castilla J, Saa P, Hetz C, Soto C. In vitro generation of infectious scrapie prions. Cell 2005;121:195-206.
- [103] Miele G, Manson J, Clinton M. A novel crythroid-specific marker of transmissible spongiform encephalopathies. Nat Med 2001;7: 361-4.
- [104] Glock B, Winter M, Rennhofer SO, Brunholzl E, Troscher D, Reisacher RB, et al. Transcript level of erythroid differentiation-related factor, a candidate surrogate marker for transmissible spongiform encephalopathy diseases in blood, shows a broad range of variation in healthy individuals. Transfusion 2003;43:1706-10.
- [105] Dietz K, Raddatz G, Wallis J, Müller N, Zerr I, Lefèvre H, et al. How much would the exclusion of transfusion recipients from donating blood reduce the spread of vCJD? Emerging Infectious Dis 2007;13:89-96.
- [106] Arbeitskreis Blut (National Advisory Committee 'Blood'). Testing plasma donations for hepatitis B core antigen (anti-HBc) in order to improve safety of cellular blood components and of quarantined fresh frozen plasma. (Recommendation 31). Bundesgesundheitsbl Gesundheitsforsch Gesundheitsschutz 2005;48:698-9. http://www.rki.de/cln_011/nn_231708/EN/Content/Prevention/NAC_Blood/V_31E.templateId=raw.property=publicationFile.pdf/V_31E.